

Inhibition of the Growth-Related Fiber Type Shift in Rat Soleus Muscle by Hypobaric-Hypoxia

Akihiko ISHIHARA¹, Kazuo ITOH², Minoru ITOH³, Chiyoko HIROFUJI⁴ and Hitomi HAYASHI¹

¹Laboratory of Neurochemistry, Faculty of Integrated Human Studies, Kyoto University, Kyoto; ²Graduate School of Integrated Science and Art, University of East Asia, Shimonoseki; ³Department of Exercise Physiology, Tenri University, Tenri; ⁴Department of Exercise Physiology, Osaka Gakuin University, Suita, Japan

Received May 13 1996; accepted August 23 1996

Summary. The histochemical type shift of fibers from fast-twitch oxidative glycolytic (FOG) to slow-twitch oxidative (SO) in the rat soleus muscle occurs during postnatal development. Our recent experiments demonstrated that the type shift of fibers from FOG to SO is inhibited by hypobaric-hypoxia. Furthermore, the present study has shown that hypobaric-hypoxia inhibits the growth-related type shift of fibers from FOG to SO in the rat soleus muscle, irrespective of the duration of exposure to hypobaric-hypoxia.

Key words—histochemistry, hypobaric-hypoxia, muscle fiber type, rat, soleus muscle.

INTRODUCTION

Mammalian skeletal muscle fibers are classified into fast-twitch glycolytic (FG), fast-twitch oxidative glycolytic (FOG), or slow-twitch oxidative (SO), according to their enzyme histochemical profiles.¹²⁾ The type shift of fibers from FOG to SO in the rat soleus muscle occurs during postnatal development.^{10,16)} Previously, we found that the growth-related type shift of fibers from FOG to SO in the rat soleus muscle was inhibited by hypobaric-hypoxia.^{5,8)} In addition, we have shown that this inhibition was induced by hypobaric-hypoxia for 5 weeks, irrespective of the age at which animals were exposed to hypobaric-hypoxia.⁶⁾

In the present study, to determine whether hypoxic-exposure for more than 5 weeks produces similar results, the change in the fiber type distribution of the

rat soleus muscle was examined after 10 weeks of hypobaric-hypoxia of 463 torr, equivalent to an altitude of 4,000 m.

MATERIALS AND METHODS

Animals and hypoxic-exposure

Ten male Sprague-Dawley rats were randomly divided into two hypoxic groups of five animals each. The animals in the hypoxic groups were housed in a hypobaric chamber (Yamashita Technology Systems, Tokushima, Japan) and from the postnatal age of 5 or 10 weeks were exposed to hypobaric-hypoxia of 463 torr, equivalent to an altitude of 4,000 m, for 10 weeks. The pressure in the chamber was returned to sea level atmospheric pressure twice a week, during which time food and water were replaced and the chamber was cleaned. The air flow in the chamber was regulated at 20 l·min⁻¹. Four normoxic groups of six male rats each were housed at sea level atmospheric pressure and used as controls at 5, 10, 15, and 20 weeks of age. All the animals were housed in a controlled environment with 12 h of daylight 12 h of darkness and maintained in room air at a temperature of 22±2°C. Standard food and water were provided ad libitum.

Histochemistry

The animals were anesthetized with sodium pentobarbital (50 mg/kg, i.p.). After being weighed, the soleus muscle was removed and weighed while moist. The muscle was immediately frozen in isopentane cooled in a mixture of dry ice and acetone. Serial

Correspondence: Akihiko Ishihara, Ph.D., Laboratory of Neurochemistry, Faculty of Integrated Human Studies, Kyoto University, Sakyo-ku, Kyoto 606-01, Japan.

Table 1. Body weights and soleus muscle weights of normoxic and hypoxic rats

	n	Body weight	Muscle weight	
		(g)	(mg)	(mg/100 g BW)
5 wks normoxia	6	156.1 ± 7.0	61.5 ± 4.8	39.6 ± 4.6
10 wks normoxia	6	365.1 ± 16.0	148.4 ± 12.2	40.7 ± 3.5
15 wks normoxia	6	479.1 ± 21.8	175.5 ± 12.8	36.7 ± 2.9
15 wks hypoxia	5	392.6 ± 18.9**	172.1 ± 13.4	43.8 ± 2.9*
20 wks normoxia	6	546.8 ± 31.7	194.8 ± 16.2	35.7 ± 2.7
20 wks hypoxia	5	403.7 ± 24.4**	177.1 ± 20.0	43.9 ± 4.7*

Values are means ± standard deviations. n, number of animals analyzed; mg/100 g BW, mg per 100 g body weight. *p < 0.01, **p < 0.001 compared with the value in the normoxic group at the corresponding age.

transverse sections, 10- μ m thick, of the widest portion of the muscle belly were cut in a cryostat maintained at -20°C . The sections were stained for adenosine triphosphatase following alkaline preincubation at pH 10.3, succinate dehydrogenase, and α -glycerophosphate dehydrogenase activities. The soleus muscle fibers were classified into FOG, SO, or intermediate (INT) as described previously.³⁾ The muscle fiber type distribution was calculated by counting the number of each type of muscle fiber in the entire transverse sections of the muscle. The cross-sectional areas of each type of muscle fiber were measured on the sections stained for adenosine triphosphatase activity, using a digitizer connected to a computer. For each muscle sample, more than 100 fibers were measured and at least 50 fibers of FOG or SO measured.

Statistics

Means and standard deviations were calculated from individual values using standard procedures. Values were represented as means \pm standard deviations. A one-way analysis of variance was used to determine any significant differences among the groups.

RESULTS

Body weight and muscle weight

The body weight was lower in the hypoxic group than in the normoxic group at 15 and 20 weeks (Table 1). There was no significant difference in the soleus muscle weight between the normoxic and hypoxic groups at 15 and 20 weeks, while the soleus muscle

Table 2. Total fiber numbers of the soleus muscle of normoxic and hypoxic rats

	n	Muscle fiber number
5 wks normoxia	6	2448 ± 236
10 wks normoxia	6	2373 ± 207
15 wks normoxia	6	2438 ± 222
15 wks hypoxia	5	2503 ± 183
20 wks normoxia	6	2531 ± 256
20 wks hypoxia	5	2344 ± 309

Values are means ± standard deviations. n, number of animals analyzed.

weight per body weight was higher in the hypoxic group than in the normoxic group at 15 and 20 weeks (Table 1).

Total fiber number

There was no significant difference in the total fiber number of the soleus muscle between the normoxic and hypoxic groups at 15 and 20 weeks (Table 2).

Fiber type distribution

In the soleus muscle, the percentage of FOG fibers was higher and that of SO fibers was lower in the hypoxic group than in the normoxic group at 15 and 20 weeks (Figs. 1 and 2). There was no significant difference in the percentage of INT fibers between the normoxic and hypoxic groups at 15 and 20 weeks (Fig. 2).

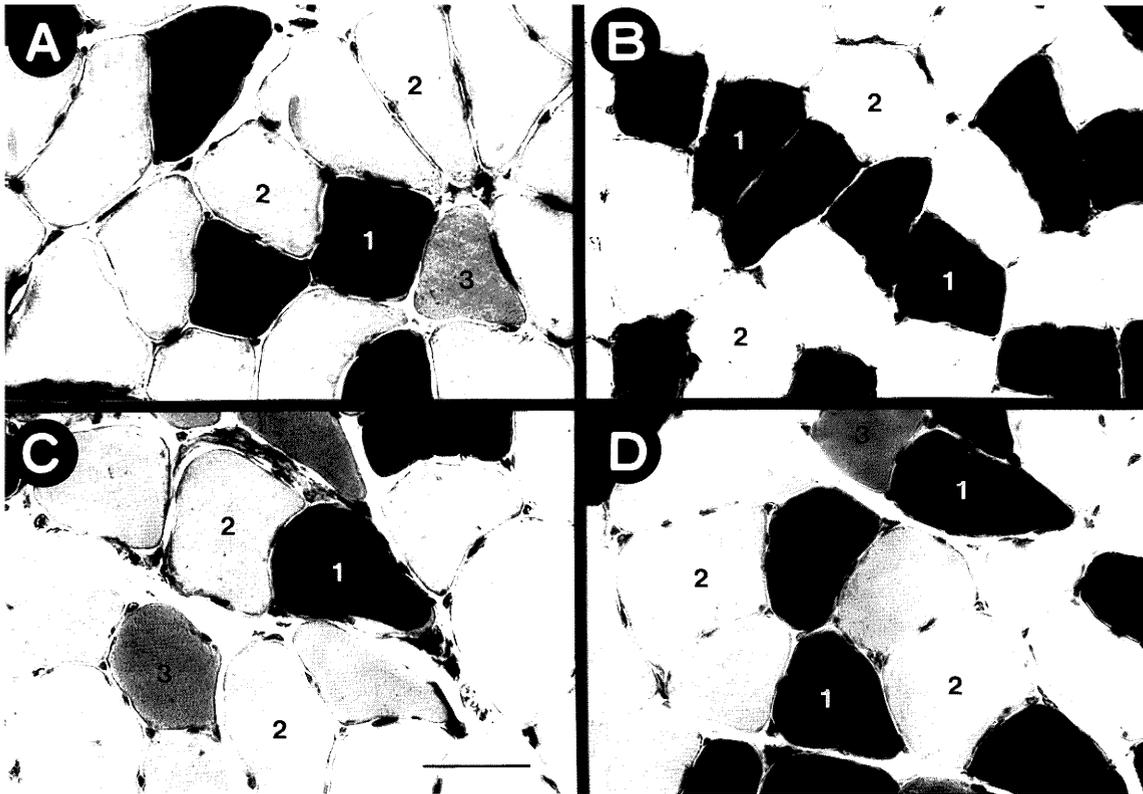


Fig. 1. Transverse sections of the soleus muscle of normoxic and hypoxic rats at 15 and 20 weeks. The sections were stained for adenosine triphosphatase activity following alkaline preincubation at pH 10.3. **A**, normoxia at 15 wks; **B**, hypoxia at 15 wks; **C**, normoxia at 20 wks; **D**, hypoxia at 20 wks; **1**, fast-twitch oxidative glycolytic; **2**, slow-twitch oxidative; **3**, intermediate. *Bar*, 50 μm for all panels.

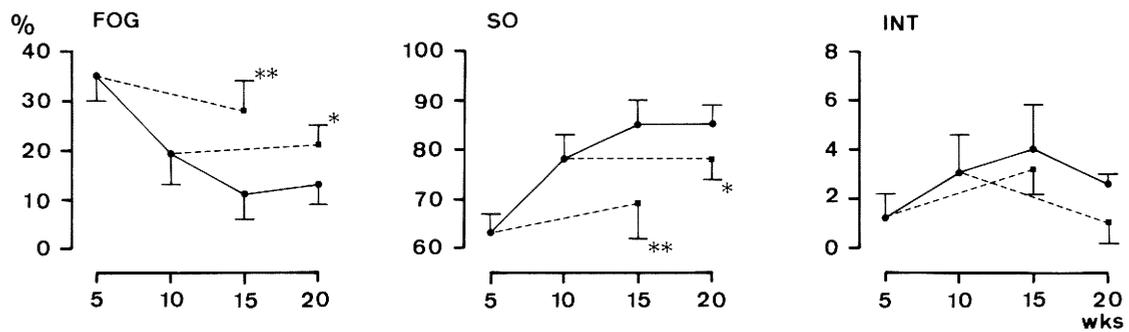


Fig. 2. Percentages of fast-twitch oxidative glycolytic (FOG), slow-twitch oxidative (SO), and intermediate (INT) fibers in the soleus muscle of normoxic (*solid lines*) and hypoxic (*dashed lines*) rats. Values are means \pm standard deviations. * $p < 0.05$, ** $p < 0.01$ compared with value in the normoxic group at the corresponding age.

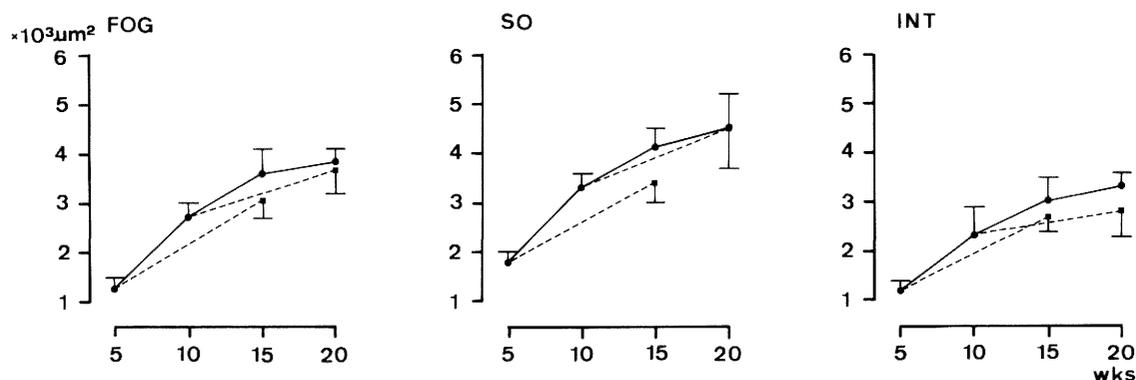


Fig. 3. Cross-sectional areas of fast-twitch oxidative glycolytic (FOG), slow-twitch oxidative (SO), and intermediate (INT) fibers in the soleus muscle of normoxic (*solid lines*) and hypoxic (*dashed lines*) rats. Values are means \pm standard deviations.

Fiber cross-sectional area

There was no significant difference in the fiber cross-sectional area of the soleus muscle between the normoxic and hypoxic groups at 15 and 20 weeks (Fig. 3).

DISCUSSION

Numerous investigations have been performed on alterations in the histochemical fiber type distribution of the skeletal muscle exposed to short- or long-term hypoxia, or chronic-hypoxia. However, data on the effect of hypoxia on muscle fiber types are contradictory because of the differences in muscles and animals and/or of hypoxic-conditions. Previous studies^{14,15} observed no changes in the fiber type distribution in the rat and guinea pig soleus muscles by exposure to hypoxia. In contrast, other studies using birds,⁹ rats,¹¹ and humans^{11,13} have found that hypoxia produces adaptative changes in the fiber type distribution in the vastus lateralis, extensor digitorum longus, tibialis anterior, and plantaris muscles.

Our previous studies^{4,8,17} have shown that hypobaric-hypoxia causes some metabolic alterations in neuromuscular systems. In the present study, the type shift of fibers from FOG to SO in the rat soleus muscle which occurs during postnatal development was inhibited for 10 weeks of hypobaric-hypoxia from 5 and 10 weeks of age. This is in agreement with our previous findings⁶ that inhibition in the

growth-related changes of histochemical fiber types and of myosin heavy chain isoforms in the rat soleus muscle was found, irrespective of the age at which the animals were exposed to 5 weeks of hypobaric-hypoxia. Therefore, we conclude that hypobaric-hypoxia inhibits the type shift of fibers from FOG to SO in the rat soleus muscle, irrespective of the age and duration of exposure to hypobaric-hypoxia.

In this study, we could not elucidate the physiological meanings of inhibition in the growth-related type shift of fibers from FOG to SO in the soleus muscle. Our previous study⁷ showed that there was no effect of hypobaric-hypoxia on functional parameters including maximal twitch force, contraction time, and fatigability of the rat soleus muscle, although hypoxic rats had a higher percentage of FOG fibers than controls. Further studies are needed to clarify the physiological meanings of inhibition in the growth-related type shift of fibers.

Male patients with chronic obstructive lung disease had a higher percentage of type II (FOG) fibers in the muscle than normal, and FOG fiber population decreased towards normal after isovolumic hemodilution.² Similarly, we found that hypoxia-acclimatized rats had a higher percentage of FOG fibers than normoxic animals, while the hypoxia-acclimatized animals showed a similar muscle fiber type distribution as found in normoxic animals after they were bred at sea level atmospheric pressure for 5 weeks.⁵ Therefore, it is suggested that a higher percentage of FOG fibers in the muscle is found only under hypoxic-conditions and is due to an acute response to hypoxia.

REFERENCES

- 1) Bigard AX, Brunet A, Guezennec CY, Monod H: Skeletal muscle changes after endurance training at high altitude. *J Appl Physiol* **71**: 2114-2121, 1991.
- 2) Hildebrand IL, Sylvén C, Esbjörnsson M, Hellström K, Jansson E: Does chronic hypoxaemia induce transformations of fibre types? *Acta Physiol Scand* **141**: 435-439, 1991.
- 3) Hirofuji C, Ishihara A, Itoh K, Itoh M, Takeuchi-Hayashi H: Fibre type composition of the soleus muscle in hypoxia-acclimatised rats. *J Anat* **181**: 327-333, 1992.
- 4) Ishihara A, Taguchi S, Itoh M, Itoh K: Oxidative metabolism of the rat soleus neuron pool following hypobaric hypoxia. *Brain Res Bull* **24**: 143-146, 1990.
- 5) Ishihara A, Itoh K, Itoh M, Hirofuji C, Hayashi H: Hypobaric-hypoxic exposure and histochemical responses of soleus muscle fibers in the rat. *Acta Histochem* **96**: 74-80, 1994.
- 6) Ishihara A, Itoh K, Oishi Y, Itoh M, Hirofuji C, Hayashi H: Effects of hypobaric hypoxia on histochemical fibre-type composition and myosin heavy chain isoform component in the rat soleus muscle. *Pflügers Arch* **429**: 601-606, 1995.
- 7) Itoh K, Moritani T, Ishida K, Hirofuji C, Taguchi S, Itoh M: Hypoxia-induced fibre type transformation in rat hindlimb muscles: histochemical and electromechanical changes. *Eur J Appl Physiol* **60**: 331-336, 1990.
- 8) Itoh M, Itoh K, Taguchi S, Hirofuji C, Takeuchi H, Ishihara A: Effect of hypobaric hypoxia on fiber type composition of the soleus muscle in the developing rat. *Aviat Space Environ Med* **63**: 583-587, 1992.
- 9) León-Velarde F, Sanchez J, Bigard AX, Brunet A, Lesty C, Monge-CC: High altitude tissue adaptation in andean coots: capillarity, fibre area, fibre type and enzymatic activities of skeletal muscle. *J Comp Physiol B* **163**: 52-58, 1993.
- 10) Maltin CA, Delday MI, Baillie AGS, Grubb DA, Garlick PJ: Fiber-type composition of nine rat muscles. I. Changes during the first year of life. *Am J Physiol* **257**: E823-E827, 1989.
- 11) Oswald O, Howald H, Di Prampero PE, Hoppeler H, Classen H, Jenni R, Bühlmann A, Ferretti G, Brückner J-C, Veicsteinas A, Gussoni M, Cerretelli P: Physiological profile of world-class high-altitude climbers. *J Appl Physiol* **60**: 1734-1742, 1986.
- 12) Peter JB, Barnard RJ, Edgerton VR, Gillespie CA, Stempel KE: Metabolic profiles of three fiber types of skeletal muscle in guinea pigs and rabbits. *Biochemistry* **11**: 2627-2633, 1972.
- 13) Rosser BWC, Hochachka PW: Metabolic capacity of muscle fibers from high-altitude natives. *Eur J Appl Physiol* **67**: 513-517, 1993.
- 14) Sillau AH, Banchemo N: Effects of hypoxia on capillary density and fiber composition in rat skeletal muscle. *Pflügers Arch* **370**: 227-232, 1977.
- 15) Sillau AH, Aquin L, Bui MV, Banchemo N: Chronic hypoxia does not affect guinea pig skeletal muscle capillarity. *Pflügers Arch* **386**: 39-45, 1980.
- 16) Smith D, Green H, Thomson J, Sharratt M: Oxidative potential in developing rat diaphragm, EDL, and soleus muscle fibers. *Am J Physiol* **254**: C661-C668, 1988.
- 17) Taguchi S, Ishihara A, Itoh M, Itoh K: Effects of hypobaric hypoxia on the oxidative capacity of the extensor digitorum longus motor units in the rat. *Neurochem Res* **15**: 923-926, 1990.