

Paradoxical Effects of Glycyrrhizin on the Induction of Granulocytes and Extrathymic T Cells between Normal Mice and Mice with Preexisting Granulocytosis

Soichiro YAMAMURA^{1,2}, Chikako MIYAJI¹, Hisami WATANABE¹, Toshihiko KAWAMURA¹, Minoru FUKUDA³ and Toru ABO¹

¹The Department of Medical Zoology and ²The Department of Orthopedics, Niigata University School of Medicine, and ³Sakamachi Hospital, Arakawa-machi, Niigata, Japan

Received July 16 1996; accepted September 2 1996

Summary. Some traditional Chinese medicines are known to have paradoxical effects. For example, one such medicine (i.e., glycyrrhizin) is effective for the treatment of allergic diseases (i.e., lymphocyte-associated inflammation), and is also sometimes effective for the treatment of suppurative diseases (i.e., granulocyte-associated inflammation). To investigate the mechanisms underlying this paradox, we conducted a study in mice. When normal mice were administered with one i. p. injection of glycyrrhizin, granulocytes and extrathymic T cells increased in proportion and number in various immune organs. However, an opposite response of glycyrrhizin was induced with such an injection in mice with preexisting granulocytosis. These mice included those treated with G-CSF and AIDS mice infected with murine leukemia virus. In other words, glycyrrhizin reduced the levels of granulocytes and extrathymic T cells in various organs of mice with preexisting granulocytosis. These results suggest that some drugs have the ability to exert paradoxical effects, depending on the conditions of the host immune system. The underlying mechanisms are discussed.

Key words—Chinese medicine, glycyrrhizin, lymphocytes, granulocytes, extrathymic T cells.

INTRODUCTION

The host defense system consists of both phagocytes and lymphocytes. The major cells of the former are granulocytes and macrophages while those of the

latter are T cells and B cells. Since granulocytes and macrophages carry a high concentration of adrenergic receptors on the surface among leukocytes, they are activated under conditions of sympathetic nerve strain, for example, anger, stress, exercise, overwork, heavy consumption of alcohol, bacterial infection, etc.¹⁻⁷⁾ Under these conditions, hosts tend to fall victim to inflammatory diseases due to the interaction of granulocytes with resident bacteria or invading bacteria.⁸⁾ On the other hand, lymphocytes express cholinergic receptors on the surface and are, therefore, activated under conditions of parasympathetic nerve dominance, for example, the absence of stress, viral infection (early phase), allergic states, etc.⁹⁻¹⁴⁾ Pulse rate is a good indicator for determining the state of the autonomic nervous system. Namely, tachycardia occurs under sympathetic nervous strain, while bradycardia is observed with parasympathetic nerve dominance.

Based on these findings and various indicators, attention has been focused on glycyrrhizin, an aqueous extract from licorice root (*Glycyrrhiza radix*).^{15,16)} This Chinese medicine is often effective for the treatment of both allergic diseases (e.g., urticaria, hypersensitivity to pollen, and atopic dermatitis) and suppurative diseases (e.g., chronic phase of viral infection, periproctal abscess, and gastric ulcer).¹⁷⁻²⁵⁾ The former diseases are caused by the overactivity of lymphocytes (accompanied by granulocytopenia), whereas the latter are caused by the overactivity of granulocytes (accompanied by lymphocytopenia). It is, therefore, possible that glycyrrhizin may have paradoxical effects even on the host defense system.

In the course of studies on glycyrrhizin, we have

Correspondence: Toru Abo, M.D., Department of Medical Zoology, Niigata University School of Medicine, Asahimachi-dori 1-757, Niigata 951, Japan.

encountered an interesting result; namely, that glycyrrhizin induces granulocytosis in normal mice while it reduces the levels of granulocytes in mice with preexisting granulocytosis. Recent studies have revealed that some oxidized substances (e.g., NO) also have some paradoxical effects as reductants or oxidants; namely, they prevent tissue damage under certain conditions while inversely damaging tissue under other conditions.²⁶⁻²⁸⁾ The present results have revealed that glycyrrhizin seems to exert similar paradoxical effects on the host defense system.

MATERIALS AND METHODS

Mice

C3H/HeN and C57BL/6 mice at the age of 5 to 8 wks, fed under specific pathogen-free conditions in our laboratory, were used for this study. All mice were originally obtained from Charles River Japan, Inc. (Atsugi, Japan).

Drugs and experimental design

Glycyrrhizin (GL), an aqueous extract of licorice root known as an anti-inflammatory substance in Chinese medicine,^{15,16)} was used. This compound consists of one molecule of glycyrrhetic acid and two molecules of glucuronic acid. In the present experiments, we used GL supplemented with glycine and cysteine (i.e., Stronger Neo-Minophagen C, SNMC). As a single high dose, SNMC was i.p. injected at a concentration of 10 mg/mouse. SNMC (2 mg/0.2 ml) was also administered intraperitoneally twice a week, as chronic low doses.

Induction of granulocytosis by G-CSF

Recombinant human G-CSF was kindly provided by Chugai Pharmaceutical Co., Tokyo, Japan. 10^4 U/mouse was i.p. injected twice a week.

Infection with LP-BM5 murine leukemia virus (MuLV)

Cell-free supernatants of LP-BM5 MuLV were prepared according to established methods by culturing G6 SC-1 clones with normal SC-1 cells for 5 days.¹⁶⁾ Mice were i.p. inoculated with 150 μ l of stock solution containing $10^{5.4} \times C$ plaque-forming units and $10^{2.7}$ mink-cell focus-forming units per milliliter. The titer of LP-BM5 MuLV components in the mixture was determined as reported previously.¹⁶⁾

Cell preparations

Mononuclear cells (MNC) were harvested from various organs of mice. Hepatic MNC were isolated by an improved method described elsewhere.²⁹⁾ Briefly, mice anesthetized with ether were sacrificed by total exsanguination via a cardiac puncture. To obtain MNC, the liver was removed, pressed through 200-gauge stainless steel mesh, and then suspended in Eagle's MEM supplemented with 5 mM HEPES (Nissui Pharmaceutical Co., Tokyo, Japan) and 2% heat-inactivated newborn calf serum. After being washed with the medium, the cell pellet was sometimes resuspended in the medium. MNC were isolated from parenchymal hepatocytes, the nuclei of hepatocytes, and Kupffer cells by Percoll (35% Percoll containing 100 U/ml heparin) gradient centrifugation.³⁰⁾

Splenocytes and thymocytes were obtained by pressing the spleen MNC and thymus through 200-gauge steel mesh; splenic erythrocytes were disrupted by 0.83% NH_4Cl -Tris buffer (pH 7.6).²⁹⁾ Bone marrow cells were obtained by flushing bilateral femurs with the medium. Blood MNC were collected by 2% dextran sedimentation for 40-50 min at room temperature, and then the supernatant was harvested. Contaminated erythrocytes were also eliminated by the NH_4Cl -Tris buffer.

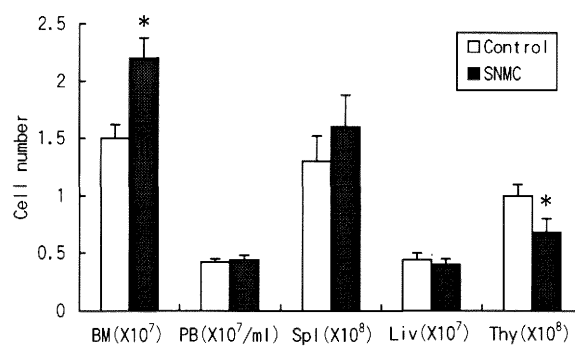
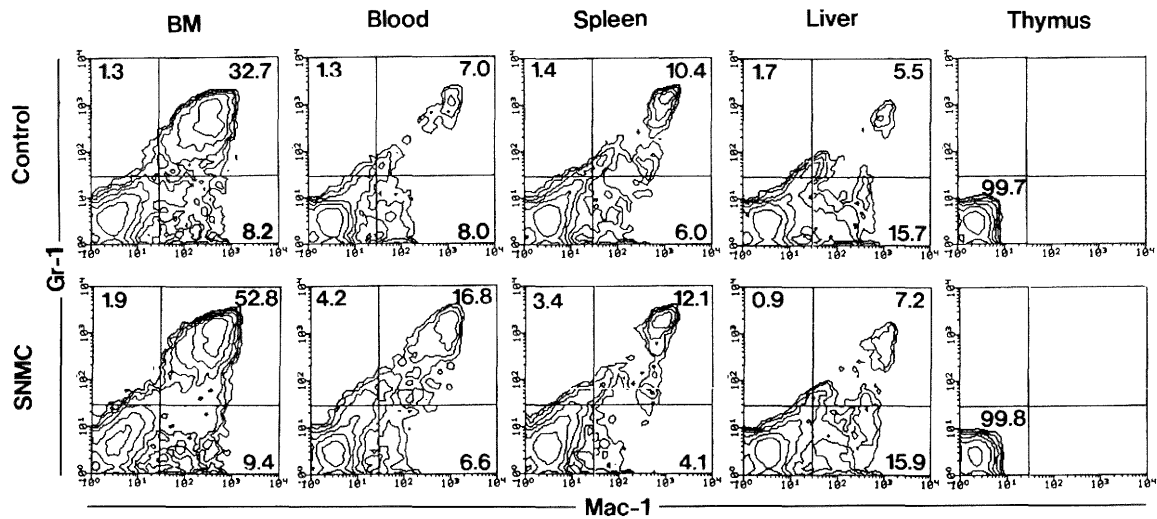
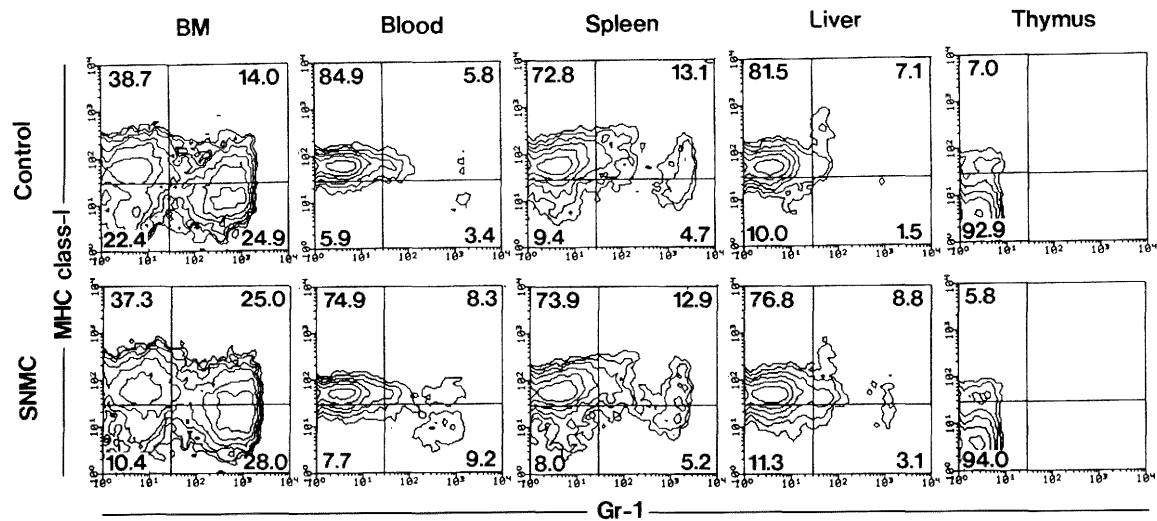


Fig. 1. Changes in the numbers of MNC in various organs of mice treated with a single injection of SNMC. SNMC (10 mg/0.2 ml) was i.p. injected and MNC were prepared on Day 3. Mean and SD of three experiments are represented. The number of MNC in the bone marrow has increased but that in the thymus decreased. * $p < 0.05$.



A



B

Fig. 2. Identification of granulocytes among MNC in various organs before and after the administration of glycyrrhizin. **A**, Two-color staining for Gr-1 and Mac-1, **B**, Two-color staining for Gr-1 and MHC class I antigens. The characterization was done on Day 3. Numbers in the figure represent the percentages of fluorescence-positive cells. Representative results of three experiments are depicted. The proportion of granulocytes has increased in the bone marrow and blood by the administration of SNMC.

Immunofluorescence test

The surface phenotypes of cells were analyzed using mAbs in conjunction with a two-color immunofluorescence test.³¹⁾ FITC, R-Phycoerythrin (R-PE), or biotin-conjugated anti-CD3 (145-2C11), anti-IL-2R β , anti-granulocyte (Gr-1, RB6-8C5), anti-erythrocyte (TER119), and anti-MHC class I (anti-mouse H-2K^b) mAbs were obtained from PharMingen Co. (San Diego, CA, U.S.A.). FITC-conjugated anti-monocyte (Mac-1) mAb (Caltag Laboratories, San Francisco, CA, U.S.A.) was also

used. Biotin-conjugated reagents were developed with PE-conjugated streptavidin (Caltag Laboratories, San Francisco, CA, U.S.A.). The fluorescence-positive cells were analyzed with a FACScan (Becton-Dickinson Co., Mountain View, CA, U.S.A.).

Statistical analysis

Statistical significance was analyzed by using Student's *t*-test.

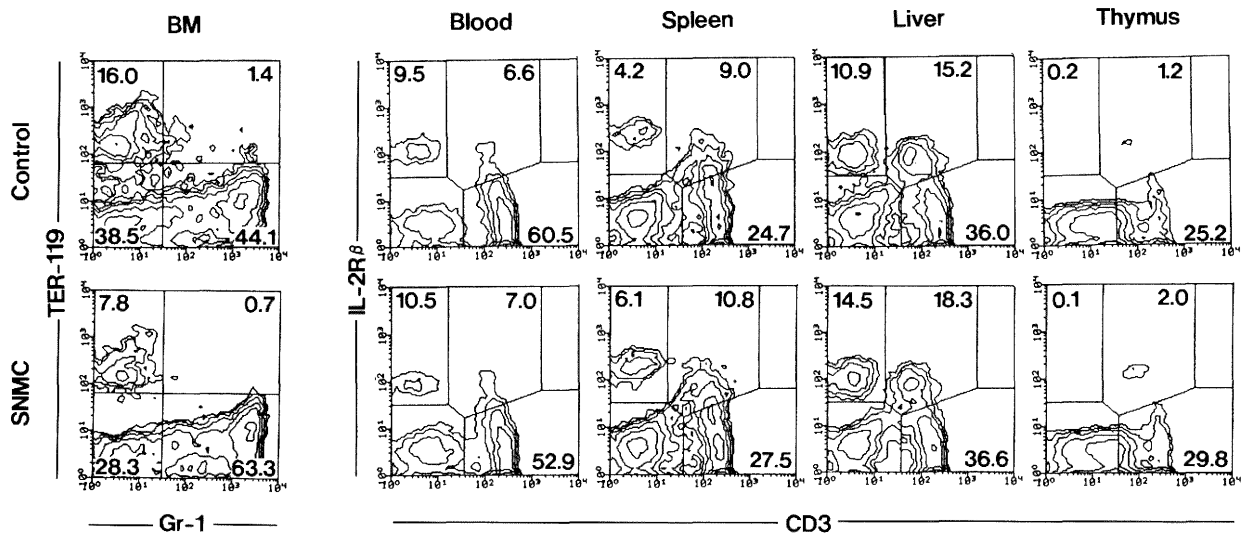


Fig. 3. Further phenotypic characterization of MNC in various organs. Two-color staining for Gr-1 and TER-119 (erythroid antigen) was performed in the bone marrow, while two-color staining for CD3 and IL-2R β was done in the other immune organs. Representative results of three experiments are depicted. The proportion of erythroid cells has decreased in the bone marrow by the administration of SNMC. IL-2R β ⁺ CD3^{int} cells tend to increase in all tested organs.

RESULTS

Effects of an acute high dose of glycyrrhizin on granulocytes

Mice were administered with a single i.p. injection of SNMC and the numbers of MNC yielded by the various immune organs were enumerated on Day 3 (Fig. 1). The number of MNC in the bone marrow increased and that in the thymus decreased ($P < 0.05$), while the numbers of MNC in other organs remained unchanged ($P > 0.05$).

To determine the variation of granulocytes, two-color staining for Gr-1 and Mac-1 and that for Gr-1 and MHC class I were performed (Fig. 2). In these stainings, granulocytes were estimated to be Mac-1⁺ Gr-1⁺ (Fig. 2A) and these granulocytes, especially in the bone marrow, were a mixture of MHC class I⁺ and class I⁻ (Fig. 2B). It was found that such granulocytes in the bone marrow, as well as in the blood, increased markedly after the injection of SNMC.

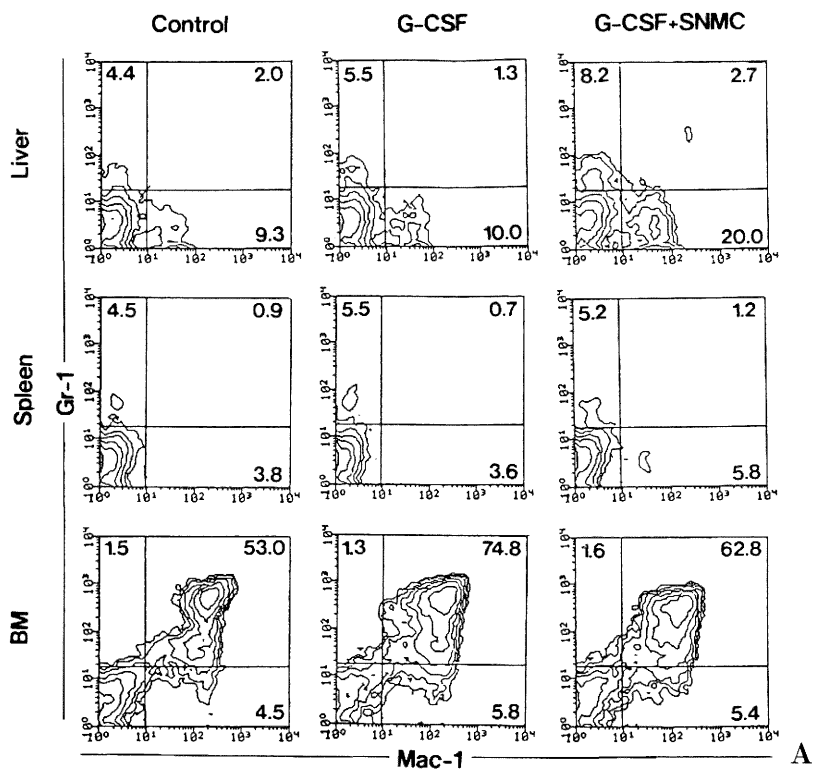
Erythroid antigen⁺ (TER-119) cells inversely decreased in the bone marrow (Fig. 3). In this figure, two-color staining for CD3 and IL-2R β was also carried out to determine the variation of IL-2R β ⁺ CD3^{int} cells (i.e., extrathymic T cells). It was confirmed that IL-2R β ⁺ CD3^{int} cells increased, especially in the liver, by the injection of SNMC. Namely, an

acute high dose of glycyrrhizin simultaneously induced an increase in the number of granulocytes in the bone marrow and an increase in the proportion of CD3^{int} cells in the liver.

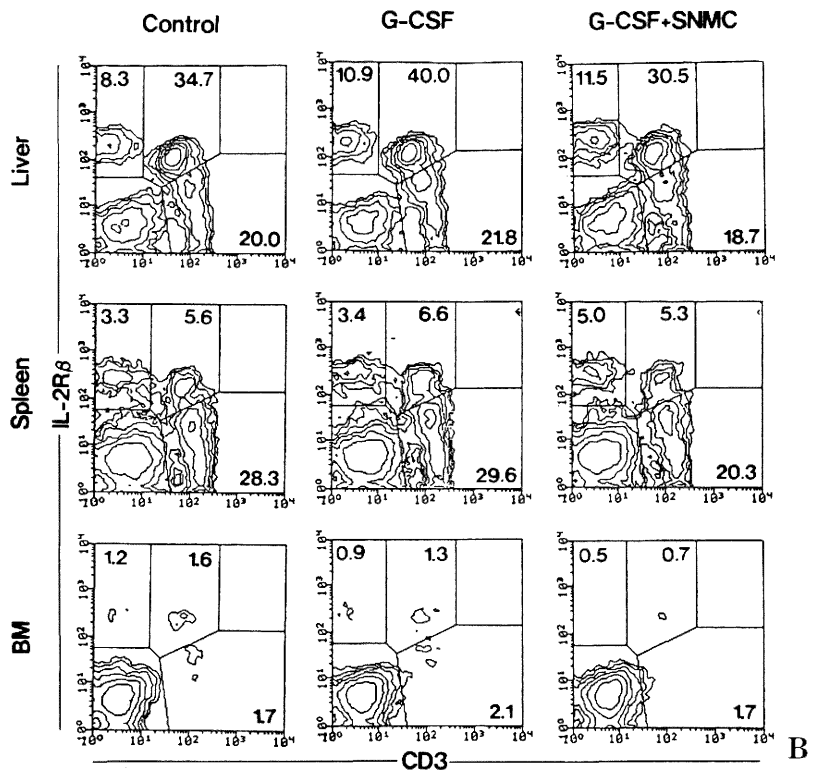
Effects of an administration of glycyrrhizin in mice with preexisting granulocytosis

In this experiment, mice were initially injected with G-CSF to induce granulocytosis. The purpose of this study was to determine whether glycyrrhizin enhances or suppresses the levels of granulocytes in these mice. In this protocol, a low-dose administration of SNMC (2 mg/one time/mouse) was repeated twice a week in conjunction with the treatment with G-CSF (also twice a week) (Fig. 4).

The number and phenotype of granulocytes were examined on Day 10. Reflecting the increase in the number of granulocytes due to the treatment with G-CSF, the number of MNC in the bone marrow increased up to 200% (data not shown). It was demonstrated that the simultaneous injection of SNMC tended to decrease the proportion of granulocytes in mice with G-CSF (75% \rightarrow 63%) in the bone marrow (Fig. 4A). Since the number of bone marrow (BM) cells in mice treated with G-CSF and SNMC also decreased (30% decrease in comparison with mice treated with G-CSF alone), the decrease in the abso-

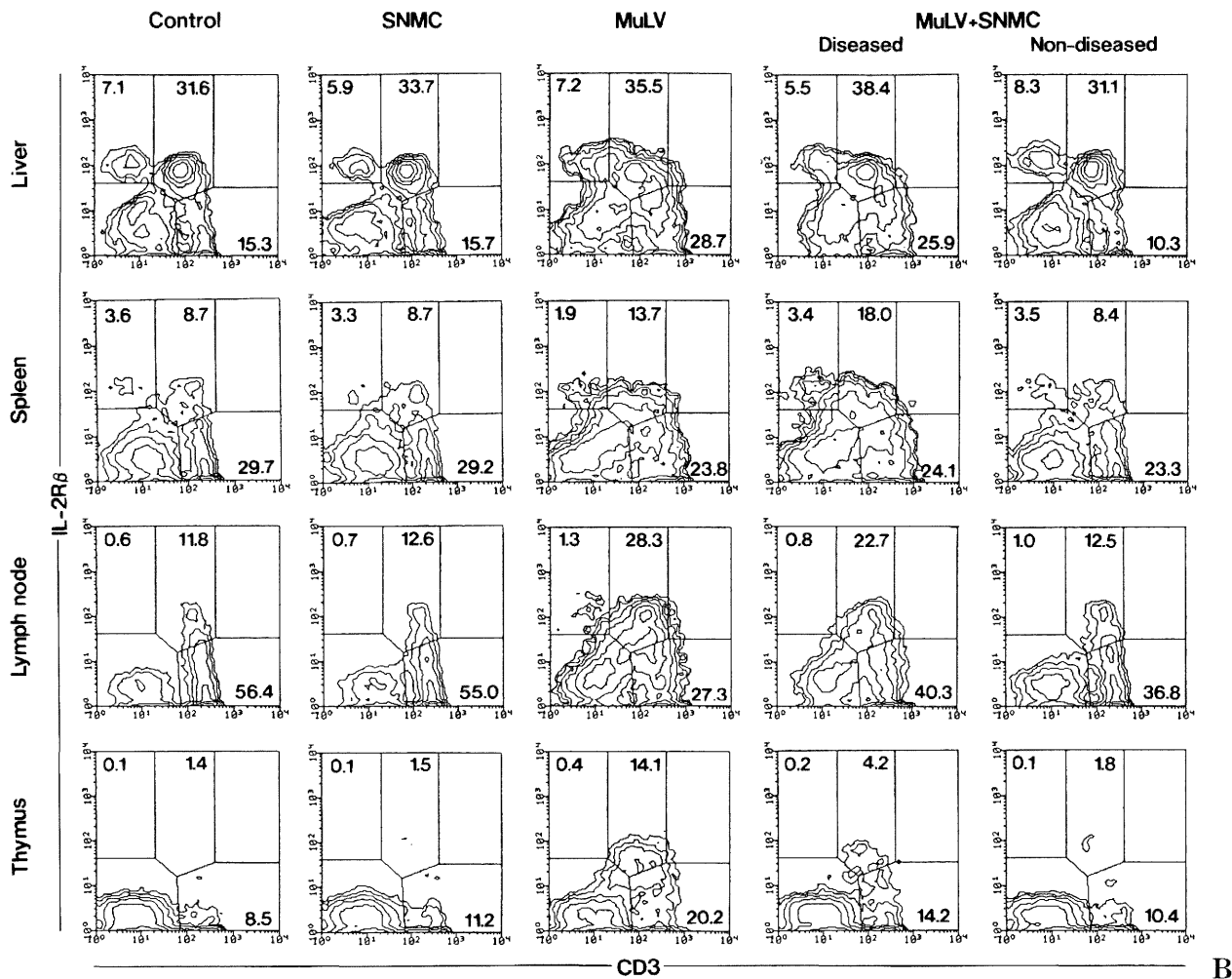


A



B

Fig. 4. SNMC suppressing the increase in the proportion of granulocytes and extrathymic T cells in mice treated with G-CSF, **A**. Two-color staining for Gr-1 and Mac-1, **B**. Two-color staining for CD3 and IL-2Rβ. Representative results of three experiments are depicted. G-CSF simultaneously increases the proportion of granulocytes and extrathymic T cells. However, such an increase is considerably suppressed by the chronic injections of a low dose of SNMC (2 mg/one time/mouse).



B

mice infected with MuLV did not show the onset of the disease (e.g., wasting syndrome) when SNMC were administered. It was demonstrated that the levels of SNMC (e.g., Mac-1⁺Gr-1⁺ cells) in the liver, spleen, and lymph nodes decreased prominently in the disease-free group of mice treated with SNMC. Control mice and mice treated with SNMC alone did not show granulocytosis.

Two-color staining for CD3 and IL-2R β is represented in Fig. 5B. As already reported, a massive expansion of CD3^{int} cells was seen in various immune organs in mice infected with MuLV. On the other hand, the disease-free group of mice treated with SNMC showed an almost normal pattern similar to that of control mice. In other words, the granulo-

cytosis and massive expansion of CD3^{int} cells were always seen as parallel events after MuLV infection.

DISCUSSION

In the present study, we demonstrated that glycyrrhizin has paradoxical effects: the increase of granulocytes and extrathymic T cells in number when injected into normal mice, and their decrease in number when injected into mice with preexisting granulocytosis. It is presumed that the present phenomenon is intimately associated with the characteristics of this Chinese medicine seen in clinical use. Namely, glycyrrhizin is used for the treatment of

allergic diseases such as urticaria, atopic dermatitis, and acute hepatitis. In these diseases, an elevated level of lymphocytes and a decreased level of granulocytes are primarily seen. On the other hand, glycyrrhizin is sometimes effective for the treatment of suppurative diseases (or granulocyte-associated diseases), including the chronic phase of viral infections (e.g., chronic active hepatitis and AIDS), peri-proctal abscess, and gastric ulcer.¹⁷⁻²²⁾ In these diseases, an elevated level of granulocytes and a decreased level of lymphocytes are inversely seen. It is concluded that glycyrrhizin has a normalization effect on the host immune system.

In answer to the question of why one substance has such contrary effects on the immune system in hosts, the following speculation is possible. Many Chinese medicines, hormones (especially steroid hormones), and NO are intermediate oxidized substances themselves and, therefore, function as both oxidants and reductants. If they function as oxidants, they simulate the sympathetic nervous system and result in granulocytosis. This is due to adrenergic receptors on granulocytes.¹⁻⁷⁾ However, if they function as reductants, they stimulate the parasympathetic nervous system and result in lymphocytosis. This is due to cholinergic receptors on lymphocytes.⁹⁻¹⁴⁾ This speculation seems to be reasonable if we consider NO. NO itself is an oxidant, stimulates the tissue function, and induces granulocytosis or tissue damage when injected or produced at a high concentration.²⁶⁻²⁸⁾ On the other hand, NO becomes a reductant (i.e., $\text{NO} \rightarrow \text{NO}_2 \rightarrow \text{NO}_3^-$), protects against tissue damage, and induces lymphocytosis when injected at chronic low doses. In the case of glycyrrhizin, if it is injected into mice with sympathetic nerve strain (i.e., granulocytosis), it tends to act as a reductant in the body. Under such conditions, many free radicals and superoxides are present (i.e., sympathetic nerve dominance). The intermediate oxidized drugs could further absorb such free radicals and superoxides.

As a rule, the levels of granulocytes and extrathymic T cells increase or decrease simultaneously. This is because both of them carry a high level of adrenergic receptors on the surface.⁸⁾ Inversely, when granulocytes and extrathymic T cells decrease, conventional T cells and B cells increase. There is a tendency for primitive leukocytes to carry a higher level of adrenergic receptors, while phylogenetically developed leukocytes carry a higher level of cholinergic receptors on their surface. It seems that their inverted regulation by autonomic nervous system may be a very important factor in the host defense system of animals and humans.

In any case, it should be understood that many

drugs, hormones, cytokines, etc., have varying characteristics as intermediate oxidative states. Therefore, depending on the conditions of the host and on the doses administered, they have the ability to function as either oxidants or reductants. This is the first report of a Chinese medicine having inverted functions in the host immune system as shown by the indicators of granulocytes and lymphocytes.

Finally, we confess that *in vitro* experiments investigating whether glycyrrhizin directly modulates the differentiation of granulocytes from their precursors remain to be done. Since there are some reports (written in Japanese) about the *in vitro* effects of glycyrrhizin on immunologic functions (e.g., PHA response), such *in vitro* experiments in the presence or absence of G-CSF should be carried out in a future study.

REFERENCES

- 1) Landmann RMA, Muller FB, Perini CH, Wesp M, Erne P, Buhler FR: Changes of immunoregulatory cells induced by psychological stress: relationship to plasma catecholamines. *Clin Exp Immunol* **58**: 127-135, 1984.
- 2) Edwards AJ, Bacon TH, Elms CA, Verardi R, Felder M, Knight SC: Changes in the populations of lymphoid cells in human peripheral blood following physical exercise. *Clin Exp Immunol* **58**: 420-427, 1984.
- 3) Ratge D, Wiedemann A, Kohse KP, Wisser H: Alterations of β -adrenoceptors on human leukocyte subsets induced by dynamic exercise: effect of prednisone. *Clin Exp Pharmacol Physiol* **15**: 43-53, 1988.
- 4) van Tits LJH, Michel MC, Grosse-Wilde H, Happel M, Eigler F-W, Soliman A, Brodde O-E: Catecholamines increase lymphocyte β_2 -adrenergic receptors via a β_2 -adrenergic, spleen-dependant process. *Amer J Physiol* **258**: E191-202, 1990.
- 5) Maisel AS, Knowlton KU, Fowler P, Rearden A, Ziegler MG, Motulsky HJ, Inset PA, Michel MC: Adrenergic control of circulating lymphocyte subpopulations. Effects of congestive heart failure, dynamic exercise, and terbutaline treatment. *J Clin Invest* **85**: 462-467, 1990.
- 6) Murray DR, Irwin M, Rearden CA, Ziegler M, Motulsky H, Maisel AS: Sympathetic and immune interactions during dynamic exercise. Mediation via a β_2 -adrenergic-dependent mechanism. *Circulation* **86**: 203-213, 1992.
- 7) Schedlowski M, Hosch W, Oberbeck R, Benschop RJ, Jacobs R, Raab H-R, Schmidt RE: Catecholamines modulate human NK cell circulation and function via spleen-independent β_2 -adrenergic mechanisms. *J Immunol* **156**: 93-99, 1996.
- 8) Fukuda M, Moroda T, Toyabe S, Iiai T, Kawachi Y, Takahashi-Iwanaga H, Iwanaga T, Okada M, Abo

- T: Granulocytosis induced by increasing sympathetic nerve activity contributes to the incidence of acute appendicitis. *Biomed Res* **17**: 171-181, 1996.
- 9) Gordon MA, Cohen JJ, Wilson IB: Muscarinic cholinergic receptors in murine lymphocytes: demonstration by direct binding. *Proc Natl Acad Sci USA* **75**: 2902-2904, 1978.
 - 10) Richman DP, Arnason BGW: Nicotinic acetylcholine receptor: Evidence for a functionally distinct receptor on human lymphocytes. *Proc Natl Acad Sci USA* **76**: 4632-4635, 1979.
 - 11) Rabey JM, Shenkman L, Gilad GM: Cholinergic muscarinic binding by human lymphocytes: changes with aging, antagonist treatment, and senile dementia of the alzheimer type. *Ann Neurol* **20**: 628-631, 1986.
 - 12) Laskowska-Bozed H, Filipowicz-Sosnowska A, Zubrzycka-Sienkiewicz Z, Ryzewski J: Expression of muscarinic cholinergic receptors on lymphocytes in various subsets of rheumatoid arthritis and their variabilities connected with treatment. *J Rheumatol* **21**: 1214-1219, 1994.
 - 13) Hara H, Hayashi K, Ohta K, Itoh N, Ohta M: Nicotinic acetylcholine receptor mRNAs in myasthenic thymuses: Association with intrathymic pathogenesis of myasthenia gravis. *Biochem Biophys Res Comm* **194**: 1269-1275, 1993.
 - 14) Wheatley LM, Urso D, Zheng Y, Loh E, Levinson AI: Molecular analysis of intrathymic nicotinic acetylcholine receptor. *Ann NY Acad Sci USA* **681**: 74-82, 1993.
 - 15) Kimura M, Watanabe H, Abo T: Selective activation of extrathymic T cells in the liver by glycyrrhizin. *Biotherapy* **5**: 167-176, 1992.
 - 16) Watanabe H, Miyaji C, Makino M, Abo T: Therapeutic effects of glycyrrhizin in mice infected with LP-BM5 murine retrovirus and mechanisms involved in the prevention of disease progression. *Biotherapy* **9**: 1-12, 1996.
 - 17) Baba M, Shigeta S: Antiviral activity of glycyrrhizin against varicella-zoster virus *in vitro*. *Antiviral Res* **7**: 99-107, 1987.
 - 18) Itoh M, Nakashima H, Baba M, Pauwel R, Clercq ED, Shigeta S, Yamamoto N: Inhibitory effect of glycyrrhizin on the *in vitro* infectivity and cytopathic activity of the human immunodeficiency virus [HIV(HTLV-III/LAV)]. *Antiviral Res* **7**: 127-137, 1987.
 - 19) Mori K, Ishida N, Uchida T, Kariyone S, Endo Y, Miura A: Effect of glycyrrhizin (SNMC: Stronger Neo-Minophagen C) in hemophilia patients with HIV infection. *Tohoku J Exp Med* **158**: 25-35, 1989.
 - 20) Itoh M, Sato A, Hirobayashi K, Tanabe F, Shigeta S, Baba M, Clercq ED, Nakashima H, Yamamoto N: Mechanism of inhibitory effect of glycyrrhizin on replication of human immunodeficiency virus (HIV). *Antiviral Res* **10**: 289-298, 1988.
 - 21) Mori K, Sakai H, Suzuki S, Akutsu Y, Isikawa M, Imaizumi M, Tada K, Aihara M, Sawada Y, Yokoyama M, Sato Y, Endo Y, Suzuki Z, Sato S, Sasaki H, Yokoyama S, Hayashi T, Uchida T, Hiwatashi K, Ishida N, Fujimaki M, Yamada K: Effects of glycyrrhizin (SNMC: Stronger NeoMinophagen C) in hemophilia patients with HIV-1 infection. *Tohoku J Exp Med* **162**: 183-193, 1990.
 - 22) Abe N, Ebina TY, Ishida N: Interferon induction by glycyrrhizin and glycyrrhetic acid in mice. *Microbiol Immunol* **26**: 536-539, 1982.
 - 23) Suzuki H, Ohta Y, Takino T, Fujisawa K, Hirayama C: Effects of glycyrrhizin on biochemical tests in patients with chronic hepatitis. Double blind trial. *Asian Med J* **26**: 423-438, 1983.
 - 24) Mizugaki M, Itoh K, Hayasaka M, Ishiwata S, Nozaki S, Nagata N, Hanadate K, Ishida N: Monoclonal antibody-based enzyme-linked immunosorbent assay for glycyrrhizin and its aglicon, glycyrrhetic acid. *J Immunoassay* **15**: 21-34, 1994.
 - 25) Zhang Y-H, Isobe K-I, Iwamoto T, Nakashima I: Bidirectional control by glycyrrhizin of the growth response of lymphocytes stimulated through a receptor-bypassed pathway. *Immunol Letters* **32**: 147-152, 1992.
 - 26) Dugas B, Mossalayi MD, Damais C, Kolb J-P: Nitric oxide production by human monocytes: evidence for a role of CD23. *Immunol Today* **16**: 574-580, 1995.
 - 27) Huang F-P, Feng G-J, Lindop G, Stott DI, Liew Y: The Role of interleukin 12 and nitric oxide in the development of spontaneous autoimmune disease in MRL/MP-*lpr/lpr* mice. *J Exp Med* **183**: 1447-1459, 1996.
 - 28) DiNapoli MR, Calderon CL, Lopez DM: The altered tumoricidal capacity of macrophages isolated from tumor-bearing mice is related to reduced expression of the inducible nitric oxide synthase gene. *J Exp Med* **183**: 1323-1329, 1996.
 - 29) Watanabe H, Ohtsuka K, Kimura M, Ikarashi Y, Ohmori K, Kusumi A, Ohteki T, Seki S, Abo T: Details of an isolation method for hepatic lymphocytes in mice. *J Immunol Methods* **146**: 145-154, 1992.
 - 30) Goossens PL, Jouin H, Marchal G, Milon G: Isolation and flow cytometric analysis of the free lymphomyeloid cells present in murine liver. *J Immunol Methods* **132**: 137-144, 1990.
 - 31) Iiai T, Watanabe H, Seki S, Sugiura K, Hirokawa K, Utsuyama M, Takahashi-Iwanaga H, Ohteki T, Abo T: Ontogeny and development of extrathymic T cells in mouse liver. *Immunology* **77**: 556-563, 1992.